

BAP1 loss confers ferroptosis resistance to cholangiocarcinoma via TLCD1-mediated membrane phospholipid remodeling

Yu Zhao¹, Peyton Classon¹, Danielle M. Carlson¹, Hidemi kawanishi¹, Jayla T. Millender¹, Irene K. Yan², Sumera I. Ilyas¹, Rory L. Smoot³, Gregory J. Gores¹, Tushar C. Patel², Mark Truty³, Davide Povero^{1,4}
¹Division of Gastroenterology and Hepatology, Mayo Clinic, Rochester, MN. ²Department of Hepatology and Liver Transplantation, Mayo Clinic, Jacksonville, FL. ³Division of Hepatobiliary and Pancreatic Surgery, Mayo Clinic, Rochester, MN. ⁴Department of Biochemistry and Molecular Biology, Mayo Clinic, Rochester, MN. Contact: povero.davide@mayo.edu

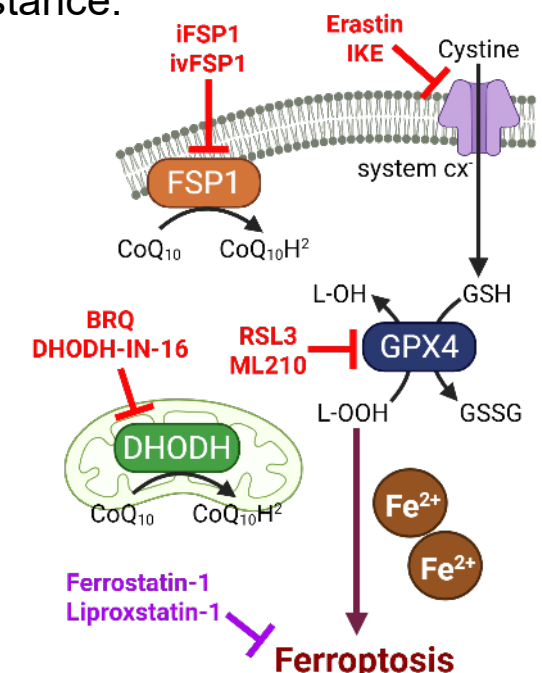
BACKGROUND

Cholangiocarcinoma (CCA) is a highly lethal epithelial cell malignancy of the biliary tract. CCA is characterized by a desmoplastic and immunosuppressive tumor microenvironment. CCA overall survival is <10% and only limited treatment options are available. Incidence and mortality are rising worldwide.

BAP1 (Ubiquitin C-terminal hydrolase deubiquitinase)

- Tumor suppressor and deubiquitinase
- Mutated in up to 32% of human CCAs
- Nuclear epigenetic regulator (H2AK119ub1 DUB)
- Cytosolic regulator cellular metabolism and cell death

Ferroptosis is an iron-catalyzed cell death driven by phospholipid peroxidation. Cancer metabolic reprogramming and genetic aberration may confer ferroptosis resistance.



AIM

The goal of this study is to identify how BAP1 loss promotes cell death evasion through metabolic reprogramming. This work may identify novel therapeutic opportunities for BAP1-mutant CCA.

METHODS

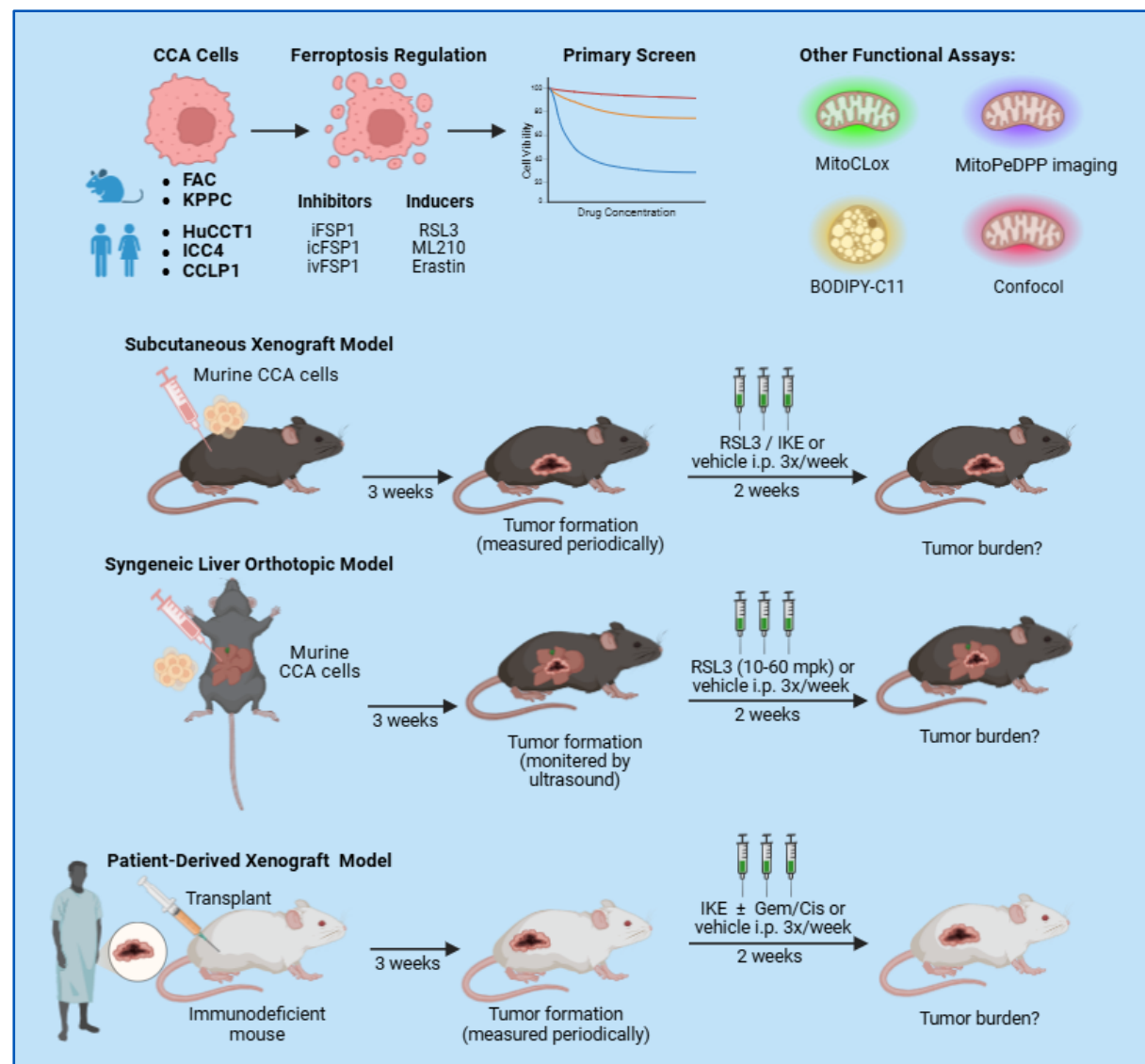


Figure 1. *In vitro* drug screening identified ferroptosis-sensitive and ferroptosis resistant CCA cells.

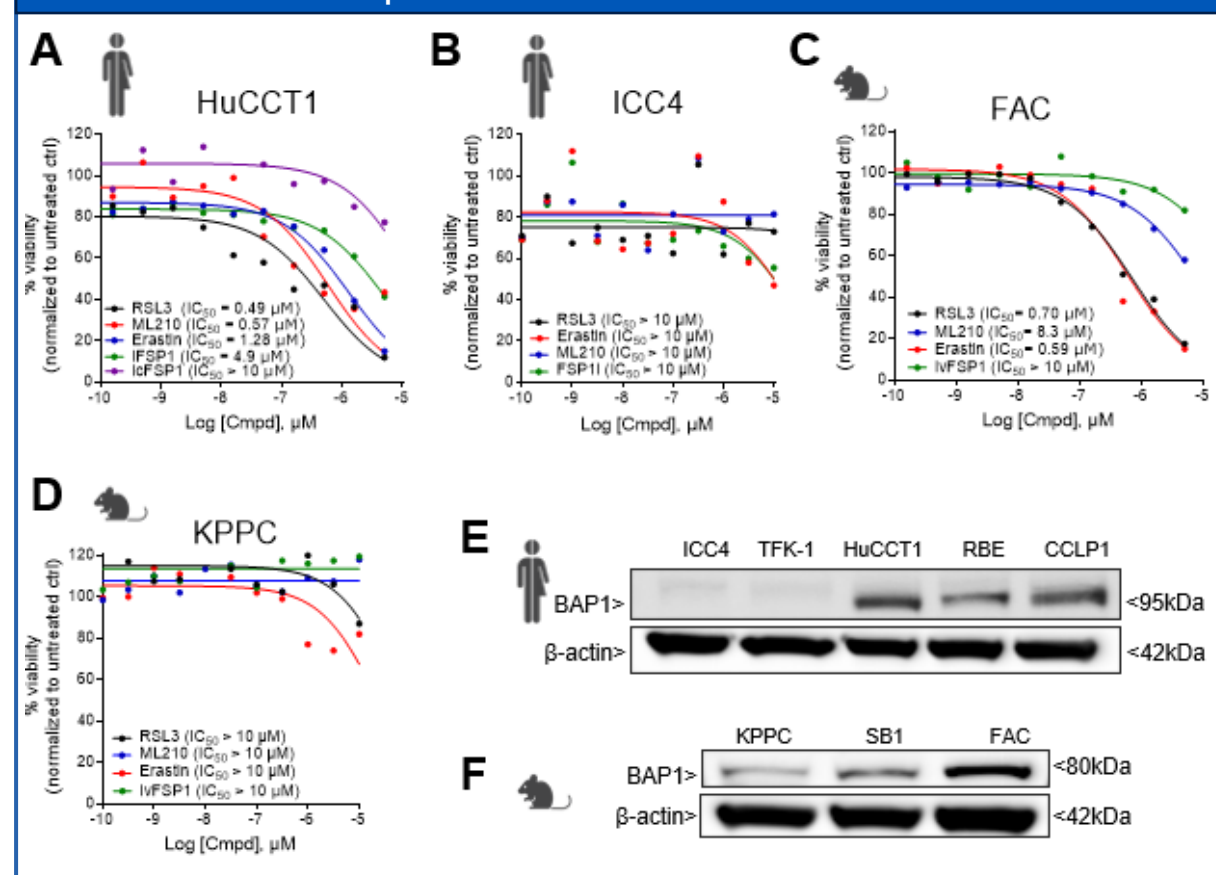


Figure 1. (A–C) Dose–response curves of human CCA cell lines (HuCCT1 and ICC4) treated with ferroptosis inducers. (D–E) Dose–response curves of murine CCA cell lines (FAC and KPPC) treated with ferroptosis inducers. Immunoblot analysis of BAP1 protein expression in (E) human CCA cell lines and (F) murine CCA cell lines. β -actin was used as loading control.

Figure 2. BAP1 loss confers CCA resistance to ferroptosis

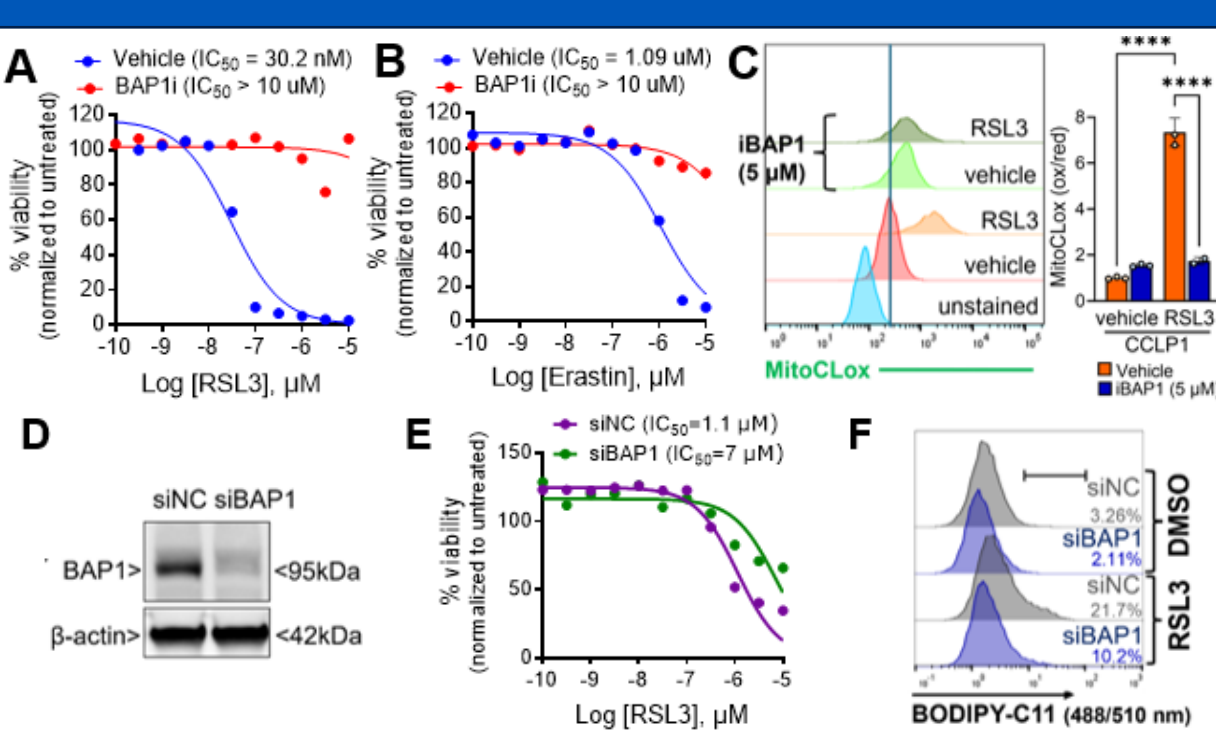


Figure 2. (A–B) Dose–response curves of RSL3 and Erastin in CCLP1 cells treated \pm BAP1 inhibitor (1 μ M). (C) Flow cytometry for mitochondrial lipid peroxidation probe MitoCLOx in CCLP1 cells \pm RSL3 and BAP1 inhibitor. (D) Immunoblot of BAP1 in siNC vs. siBAP1 cells. (E) Dose–response curve of RSL3 in siNC vs. siBAP1 CCLP1 cells. (F) Flow cytometry of C11-BODIPY lipid peroxidation assay in siNC and siBAP1 cells \pm RSL3.

Figure 3. BAP1 loss protects CCA from mitochondrial lipid peroxidation

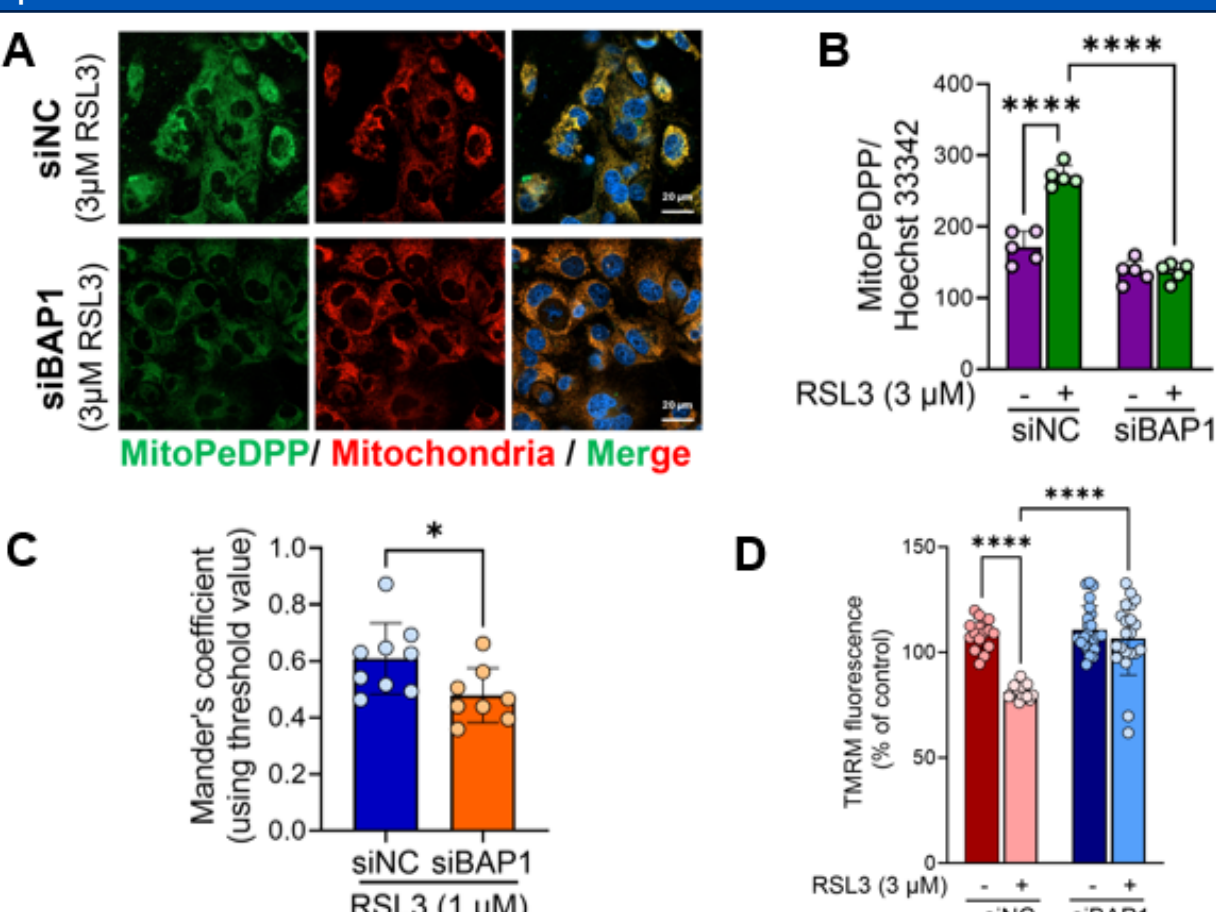


Figure 3. (A–B) Representative confocal images and quantification of mitochondrial lipid peroxidation probe MitoPeDPP (green) and mitochondria (red) in CCLP1 \pm RSL3. (C) Colocalization of C11-BODIPY and Mitotracker assessed by Mander's coefficient in siNC vs. siBAP1 CCLP1 cells. (D) Mitochondrial membrane potential measured by TMRM in siNC vs. siBAP1 CCLP1 \pm RSL3.

Figure 4. BAP1 loss remodels membrane phospholipid composition via TLCD1 activation.

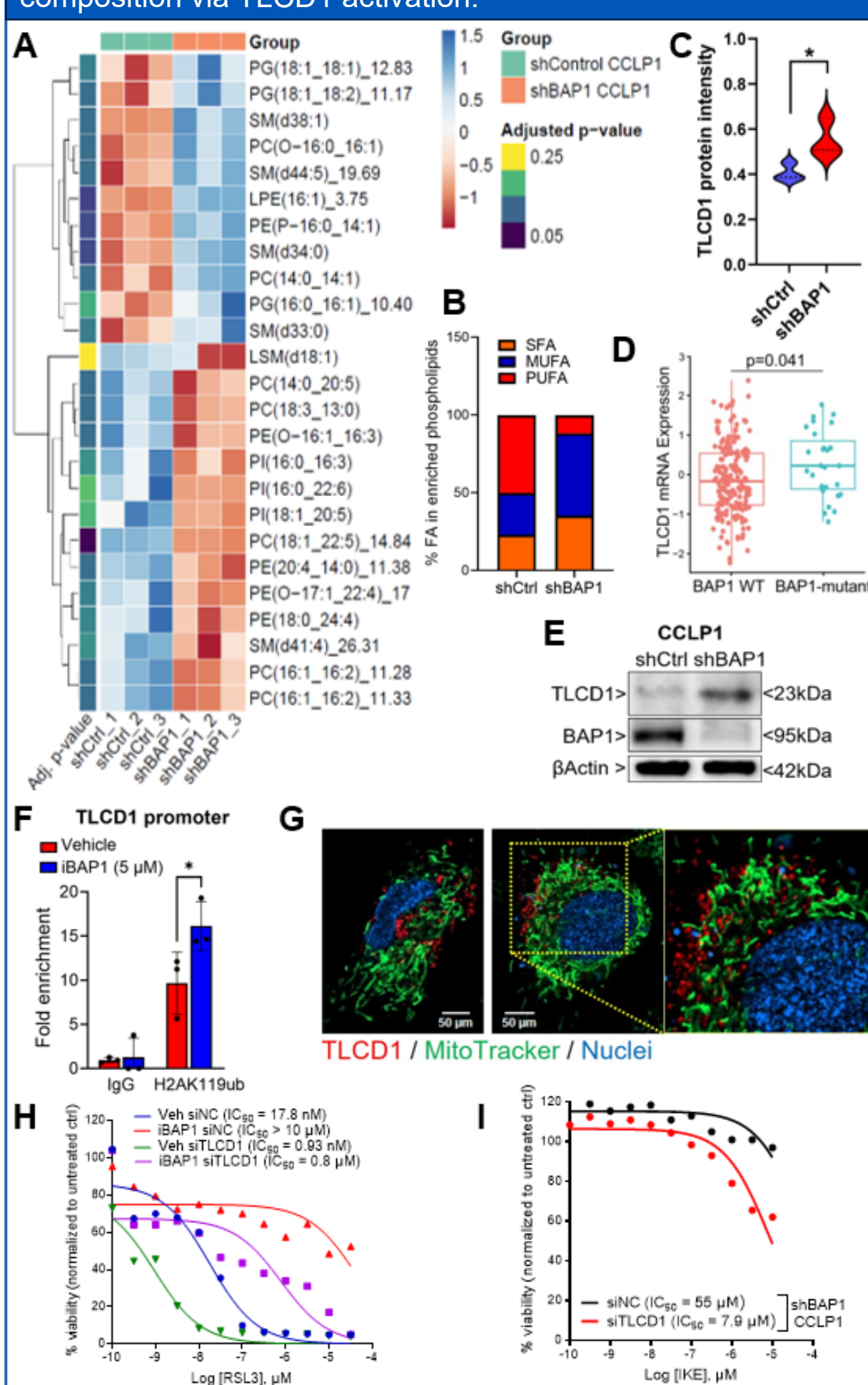


Figure 4. (A–B) Phospholipidomics heatmap and fatty acid-PL enrichment in shControl vs. shBAP1 CCLP1 cells. (C) Proteomics analysis identified lysophosphatidylethanolamine acyltransferase TLCD1 enrichment in shBAP1 CCLP1 cells. (D) TLCD1 RNA expression in WT and BAP1-mutant human CCA subjects (Mayo Clinic cohort, n=222 subjects). (E) TLCD1 immunoblot in shControl and shBAP1 CCLP1 cells. (F) ChIP–PCR for H2AK119ub1 at TLCD1 locus showing increased promoter occupancy upon BAP1 loss. (G) Confocal images of TLCD1 and Mitotracker staining. (H–I) Dose–response curves of RSL3 and IKE in BAP1-deficient CCLP1 cells \pm siTLCD1.

Figure 5. Ferroptosis reduces CCA tumor burden and induces lipid peroxidation in murine and human CCA

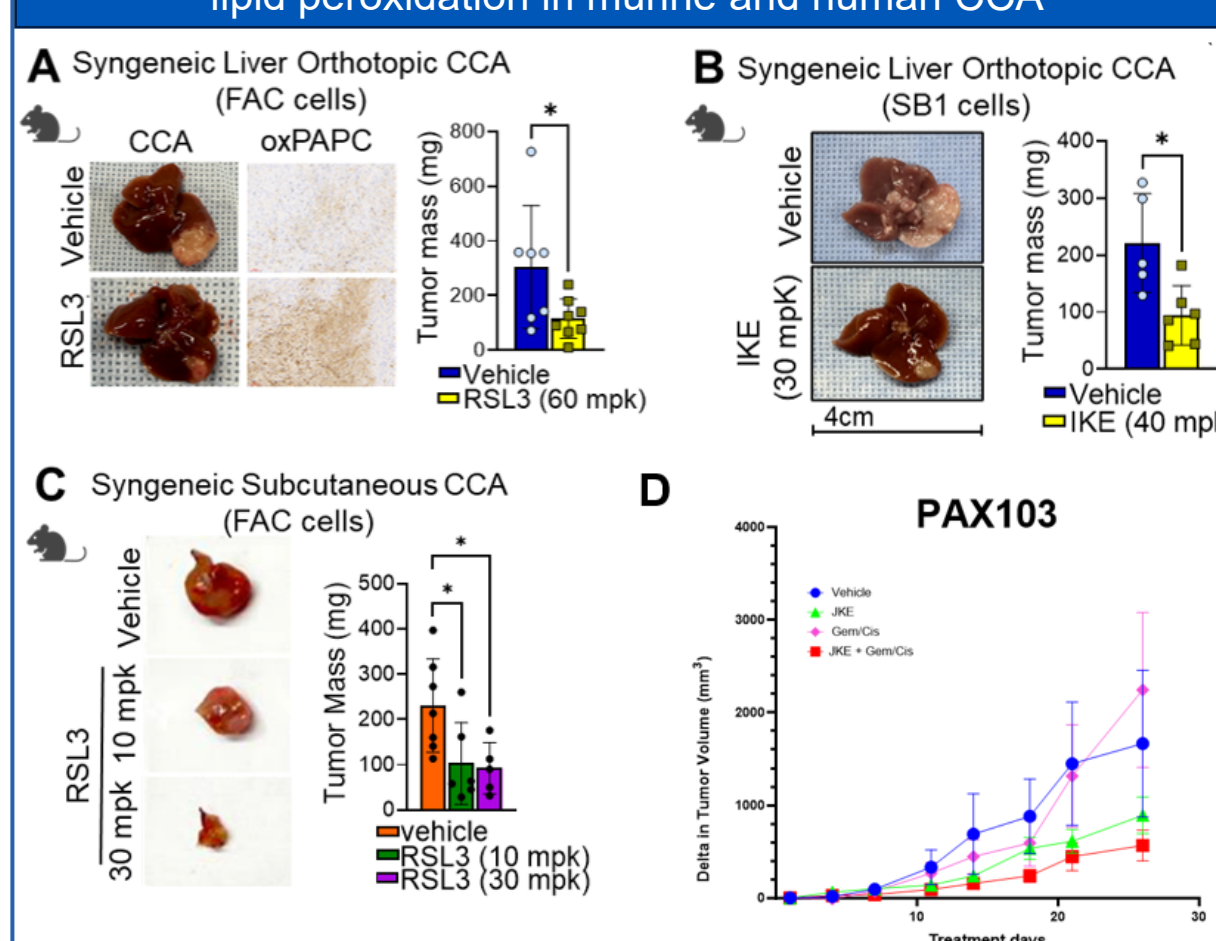


Figure 5. (A–B) Ferroptosis inducer RSL3 or IKE reduces tumor growth in two syngeneic orthotopic CCA cell liver implantation models (FAC and SB1 cells). (C) Ferroptosis suppresses tumor growth in syngeneic subcutaneous CCA models. (D) Ferroptosis inducer JKE-1674 (20 mpk) alone or with gemcitabine and cisplatin (Gem/Cis) reduces tumor burden of CCA patient-derived xenografts (PDXs). Ferroptosis inducers were administered intraperitoneally, intratumor or orally. mpk=mg/Kg.

CONCLUSIONS:

- BAP1 loss confers CCA resistance to ferroptosis, especially via protection against mitochondrial lipid peroxidation.
- BAP1 loss remodels membrane phospholipid composition via TLCD1 by enriching for ferroptosis-blocking MUFA-PLs at the expense of ferroptosis-inducing PUFA-PLs.
- *In vivo* ferroptosis reduces tumor growth in tumor-bearing murine CCA models.
- Ferroptosis inducer JKE-1674 blocks tumor growth and augments response to standard-of-care chemotherapy (Gem/Cis) in CCA PDX resistant to Gem/Cis.

ACKNOWLEDGMENTS:

- This project is supported by the:
- Cholangiocarcinoma Foundation Stewart Mather Memorial Research Fellowship
 - AASLD Pinnacle Liver Research Award